

# Nomenclatural changes, reinstatements, new combinations, and new synonymies among American Cerambycids (Coleoptera)

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**Abstract:** New synonyms: *Protorma scabrosa* Waterhouse, 1880 and *Protorma recurvatum* Williams, 1829 = *Strongylaspis costifer* Thomson, 1877; *Callopsisma ruficollis* Bates, 1870 = *Eriphus croceicollis* White, 1855; *Odontocera cinctiventris* Bates, 1870 = *Odontocera simplex* White, 1855; *Tomopterus aurantiacosignatus* Zajciw, 1969 = *Tomopterus similis* Fisher, 1930; *Cerambyx spectabilis* Voet, 1778 = *Cerambyx velutinus* Fabricius, 1775; *Chion ochraceus* Bates, 1885 = *Lamia spinifera* Fabricius, 1792; *Lissonotus shepherdi* Pascoe, 1859 = *Callidium equestre* Fabricius, 1787; *Cerambyx pulverulentus* Olivier, 1790 = *Cerambyx farinosus* Linné, 1758; *Lamia crypta* Say, 1832 = *Saperda annulata* Fabricius, 1801; *Ptericoptus forsteri* Tippmann, 1960 = *Bisaltis buquetii* Thomson, 1868; *Ataxia flaviceps* Breuning, 1942 = *Stenocorus obscurus* Fabricius, 1801; *Cryptocranium gounellei* Breuning, 1980 = *Cryptocranium cazieri* Lane, 1958; *Epectasis grossepunctata* Breuning, 1942 = *Saperda juncea* Newman, 1840; *Paraclitemnestra gigantea* Breuning, 1974 = *Jamesia lineata* Fisher, 1926; *Hippopsis tremata* Galileo and Martins, 1988 = *Hippopsis meinerti* Aurivillius, 1900; *Lamia punctata* Fabricius, 1792 = *Cerambyx (Lamia) Daviesii* Swederus, 1787; *Leiopus (Oedopeza) pogonocheroides* Audinet-Serville, 1835 = *Lamia ocellator* Fabricius, 1801; *Colobothea velutina* Bates, 1865 = *Lamia macularis* Olivier, 1792; *Xylergates dorotheae* Gilmour, 1962 = *Xylergates elaineae* Gilmour, 1962; *Colobothea velutina* Bates, 1865 = *Lamia macularis* Olivier, 1792; *Pretilia telephoroides* Bates, 1866 = *Saperda tuberculata* Fabricius, 1801; *Guyanestola macrophthalma* Breuning, 1961 = *Estoloides (Estoloides) angustifrons* Breuning, 1943; *Drycothaea marmorata* Martins and Galileo, 1990 = *Guyanestola brasiliensis* Breuning, 1974.

The following species are reinstated: *Strongylaspis costifer* Thomson, 1877; *Oxymerus lineatus* Dupont, 1836; *Hebestola operaria* Erichson, 1848; *Hesycha lateralis* Thomson, 1868.

The genus *Hephialtis* Thomson, 1864 is reinstated.

New combinations: *Hephialtis ruber* (Thunberg, 1822); *Protorma costifera* (Thomson, 1877); *Knulliana cincta spinifera* (Fabricius, 1801) comb. nov.; *Ataxia operaria* (Erichson, 1848) comb. nov.; *Paraclitemnestra lineata* (Fisher, 1926) comb. nov.; *Oedopeza ocellator* (Fabricius, 1801) comb. nov.; *Toronaeus incisus* (Bates, 1864) comb. nov.; *Pretilia tuberculata* (Fabricius, 1801) comb. nov. *Drycothaea turrialbae* (Breuning, 1943) comb. nov. *Drycothaea truncatipennis* new name for *Estola stictica* Breuning, 1942 (nec Bates, 1881)

Some host-plants are given.

## Introduction

During my various visits through several institutions sheltering historical collections, I noticed some unknown synonymies which are interesting to publish. The acronyms used are:

MNH for the Muséum d'Histoire Naturelle de Paris,  
BMNH for the British Natural History Museum in  
London,

AMNH for the American Museum of Natural History in  
New York,

USNM for the United States National Museum in Wash-  
ington D.C. (Smithsonian Institution),

ICCM for the Carnegie Museum in Pittsburgh, Pennsylv-  
ania,

MNHU for the Museum für Naturkunde der Humboldt  
Universität in Berlin,

ZMK for the Zoologisk Museum in Copenhagen,

MNRJ for the Museu Nacional da Quinta da Boa Vista  
in Rio de Janeiro,

ZSMA for the Zoologische Staatssammlung des Bay-  
erischen Staates in Munich,

NRS for the Naturhistoriska Riksmuseet in Stockholm.

Concerning host plant records, the scientific name is followed by the herbarium collection refer-

ence number sampled by a botanist; several parts are distributed throughout main botanical institutions of the world.

## 1. Reinstated species:

### Subfamily Prioninae

#### Tribe Macrotomini

***Protorma costifera* (Thomson, 1877), reinstated and new combination.**

*Strongylaspis costifer* Thomson, 1877: 275

*Protorma scabrosa* Waterhouse, 1880: 289, **new syno-  
nymy**

*Stenodontes (Protorma) scabrosus*; Lameere, 1903: 215

*Protorma recurvatum* Williams, 1929: 143, pl. V, fig. 1,  
**new synonymy**

*Protorma recurvata* Blackwelder, 1946: 552 [Catalog]

Lameere, without having seen Thomson's material, simply synonymized *Strongylaspis costifer* with *Strongylaspis corticarius* (Erichson, 1848). All the following authors (Villiers, 1980: 143 and

others) accepted Lameere's judgement. Fortunately, Thomson's type of *Strongylaspis costifer* is visible in the MNHN, and in fact, belongs to the genus *Protorma*. Later on, I saw Williams' type in ICCM of *Protorma recurvatum* and came to the conclusion that Williams species is just a synonym of Thomson's species. Williams wrote "It was carefully compared by me with the type of *Protorma scabrosa* Waterhouse, from which it is quite distinct." In fact, the description of *Protorma recurvatum* was based on a single male specimen, and the type of *Protorma scabrosa* in the BMNH is a female. The three described taxa belong to the same species, and the valid name is *Protorma costifera* (Thomson, 1877).

### Subfamily Cerambycinae

#### Tribe Trachyderini

*Oxymerus lineatus* Dupont, 1838 reinstated

*Oxymerus lineatus* Dupont, 1838: 41, pl. 211, fig. 1 [Brazil]

*Oxymerus approximatus* Dupont, 1838: 44, pl. 212, fig. 2 [Cayenne]

*Oxymerus elongatus* Dupont, 1838, 1838: 45, pl. 213, fig. 1 [Cayenne]

*Oxymerus distinguendus* Dupont, 1838: 48, pl. 214, fig. 2 [Demerary]

*Oxymerus aculeatus* subsp. *lineatus* Hüdepohl, 1979: 20, pl. I, 1(m)

*Oxymerus aculeatus* var. *lineatus* Rémy, 1988: 130

From November 1982 to January 1983 the species was particularly abundant around Kourou where I saw more than a thousand. I definitely do not agree with Hüdepohl synonymies and status changing (Hüdepohl, 1979: 20). The correct synonymy, after checking the types found in the MNHN, is listed above.

### Subfamily Lamiinae

#### Tribe Pteropliini

Fabricius described (1775: 180) a *Stenocorus annulatus* from "America meridionali" belonging to the Banks collection (today in the BMNH). Fabricius later on (1801: 310) mentioned the synonymy with *Cerambyx annulatus* Olivier, 1792 and *Cerambyx hirtipes* Degeer, 1775 (belonging to the actual genus *Colobothea* Lepeletier and Audinet-Serville, 1825). In 1792: 314 he described a *Saperda lineata* from America meridionali. The two syntypes in the ZMK belong to Degeer's species, *Cerambyx hirtipes*.

In 1801: 326, he described *Saperda annulata* and gave the short following description in latin: thorace rotundato, subspinoso, antennarum articulis basi albis, which agrees with the type in Fabricius' collection in the ZMK (conspecific with Say's *Lamia crypta*); both the locality (America meridionali) and the synonymy (*Saperda lineata* 1792: 314) were wrongly associated with the above description. The confusion was obviously caused by an error in the printing of page 326 in Fabricius' Systema Eleutheratorum. It is clear that between the *Saperda nigricornis* (n° 47) and the *Saperda tristis* (n° 51) the synonymies were wrongly skipped. Bates in the supplement of Biologia Centrali Americana (1885: 347) wrote: *Ataxia Crypta* ... Said by Erichson to be the *Saperda lineata* of Fabricius (Ent. Syst. ii p. 314), and so registered in the Munich catalogue, but it does not agree with the description, which refers also, as Fabricius states, to a South-American (Brazilian ?) insect in Lunds collection. Erichson might have seen the type and Bates read the wrong description (Fabricius, 1792: 314). The name *Saperda annulata* being used several times by Fabricius, we decide to conserve Say's species name in order to avoid more confusion. The complete synonymy of *Ataxia crypta* is the following:

#### *Ataxia crypta* (Say, 1832)

*Saperda annulata* Fabricius, 1801 (*nec* Fabricius, 1792): 326, **new synonymy**

*Lamia crypta* Say, 1832: 5

*Amniscus ? cryptus* Haldeman, 1847: 47

*Stenosoma crypta* LeConte, 1873: 302

*Ataxia crypta* LeConte, 1873: 344

*Ataxia sordida* Haldeman, 1847: 56

*Stenosoma sordida* LeConte, 1852: 158

Consequently *Hebestola operaria* Erichson, 1848 synonymized with *Saperda annulata* Fabricius, *Parysatis nigritarsis* Thomson, 1868b, and *Parysatis flavescens* Bates, 1880 becomes a valid taxon and we can write the following synonymy:

#### *Ataxia operaria* (Erichson, 1848), reinstated

*Hebestola operaria* Erichson, 1848: 574

*Parysatis nigritarsis* Thomson, 1868b: 120

*Esthlogena operaria* Lacordaire, 1872: 600

*Ataxia lineata* Gemminger and Harold, 1873: 3100 [Catalog]

*Parysatis flavescens* Bates, 1880: 112

#### Tribe Onciderini

*Lampedusa lateralis* (Thomson, 1868) reinstated

*Hesycha lateralis* Thomson, 1868a: 63 [Cayenne]  
*Lampedusa obliquator* Dillon and Dillon (*nec* Fabricius, 1801), 1945: 114, pl. V, fig. 10

***Lampedusa obliquator* (Fabricius, 1801)**

*Lamia obliquator* Fabricius, 1801: 303  
*Hypsioma obliquator* Thomson, 1868a: 50  
*Plerodia obliquator* Aurivillius, 1923: 342 [Catalog]

The type of *Hesycha lateralis* Thomson, 1868 is different from the type of *Lamia obliquator* Fabricius, 1801. Dillon and Dillon wrongly interpreted Fabricius' species and consequently *Lampedusa lateralis* (Thomson, 1868) is reinstated. Both species were encountered in French Guiana and belong to the genus *Lampedusa* as described by Dillon and Dillon, 1945: 112. Following the nomenclatural code, *Lamia obliquator* remains the type species of the genus *Lampedusa* Dillon and Dillon, 1945.

**2. Reinstated genus:**

**Subfamily Prioninae**

**Tribe Callipogonini**

The genus *Hephialtès* Thomson, 1864 was created for *Hephialtès tricostatus* Thomson, 1864: 286 based on a female specimen from Brazil. A year later he described the male as *badium*. Both are synonyms of *Trachyderes ruber* Thunberg, 1822. Lameere (1904: 28) synonymized the genus with *Anacanthus* Audinet-Serville, 1832 which he considered as a subgenus of *Stictosomus* Audinet-Serville, 1832. Whilst reading the description of the genus *Stictosomus* (page 153) we note incompatible features with Thunberg's species «Corselet sans crénelures» and «mandibules allongées, étroites, aiguës, denticulées intérieurement». Considering the description of *Anacanthus* (page 165 of the same Audinet-Serville's paper) a name which means in Greek "without spines" we find again incompatible features («corselet mutique ... arrondi...»). Neither the genus nor the subgenus retained by Lameere is correct.

Though the type species of the 3 concerned genera (*Stictosomus semicostatus* Audinet-Serville, 1832, *Anacanthus costatus* Audinet-Serville, 1832 and *Hephialtès tricostatus* Thomson, 1864) belong obviously to the same natural subdivision of the tribe Callipogonini, I think it is unreasonable to synonymize them under the same genus.

***Hephialtès* Thomson, 1864 reinstated**

Type species: *Hephialtès tricostatus* Thomson, 1864 (= *Trachyderes ruber* Thunberg, 1822) [Brazil] by original designation.

*Hephialtès* Thomson, 1864: 285

***Hephialtès ruber* (Thunberg, 1822), comb. nov.**

*Trachyderes ruber* Thunberg, 1822: 305  
*Orthosoma badium* Dejean (nomen nudum), 1837: 342  
*Hephialtès tricostatus* Thomson, 1864: 286  
*Hephialtès badium* Thomson, 1865: 577  
*Hephialtès sulcatus* Lacordaire, 1869: 147  
*Hephialtès badius* Gemminger and Harold, 1872: 2774 [Catalog]  
*Hephialtès tricostatus* Gahan, 1895: 84  
*Stictosomus (Anacanthus) tricostatus* Lameere, 1904: 28  
*Stictosomus ruber* Lameere, 1912: 164  
*Stictosomus (Anacanthus) ruber* Lameere, 1913: 38 [Catalog]  
*Anacanthus ruber* Villiers, 1980: 152, fig. 20

**3. New synonyms and consequently new combinations:**

**Subfamily Cerambycinae**

**Tribe Rhinotragini**

***Odontocera simplex* White, 1855**

*Odontocera simplex* White, 1855: 189  
*Odontocera cinctiventris* Bates, 1870a: 321, new synonymy

White's type, in the BMNH, is represented by a male specimen and Bates' type, in the MNHN, a female, although the latter was doubtful concerning the gender of his *O. cinctiventris*. I personally obtained many individuals of both sexes by rearing them on their host plant. I was already alerted by the elytral similarity of the two supposed species before having them bred. They emerged from three different species of trees all belonging to the family of Papilionaceae: *Taralea oppositifolia* Aublet [Denis Loubry 1611]; *Dipteryx punctata* (Black) Amshoff [Denis Loubry 1797]; *Monopteryx inpaë* Rodrigues [Denis Loubry 1739].

**Tribe Callichromatini**

*Callichroma velutinum* (Fabricius, 1775)  
*Cerambyx velutinus* Fabricius, 1775: 167  
*Cerambyx spectabilis* Voet, 1778: 12, pl. X, fig. 40, new synonymy  
*Cerambyx velutinus* Olivier, 1795: 24, (67) pl. VI, fig. 41  
*Callichroma velutina* Serville, 1833: 557  
*Callichroma velutinum* White, 1853: 164

*Callichroma velutinus* Lacordaire, 1869: 16  
*Callichroma porphyrogenitum* Bates, 1870: 333  
*Callichroma (Callichroma) velutinum* Schmidt, 1924: 321

Considering that the illustration is sufficient, and that the locality is "Indes Occidentales", this leaves no doubt about the identity of this taxon. The epithet "*spectabilis*" still remains in all catalogs for obscure reasons.

### Tribe Lissonotini

#### *Lissonotus equestris* (Fabricius, 1787)

*Cerambyx biserratus* Voet, 1778: 29, pl. XXV, fig. 137 [America]  
*Callidium equestre* Fabricius, 1787: 153 [Cajennae]  
*Cerambyx unidentatus* Olivier, 1795: 20, (n° 67) pl. XIX, fig. 145  
*Cerambyx cinctus* Panzer in Voet, 1798: 2, pl. XXV, fig. 136  
*Cerambyx cingulatus* Panzer in Voet, 1798: 2, pl. XXV, fig. 137  
*Lissonotus cinctus* Dalman in Schönherr, 1817: 364 [Cayenna]  
*Lissonotus unidentatus* Dalman in Schönherr, 1817: 364  
*Lissonotus equestris* Dejean, 1821: 109  
*Lissonotus shepherdii* Pascoe, 1859: 16 [Pará], **new synonymy**  
*Lissonotus shepherdii* Blackwelder, 1946: 592 [Brazil] Catalog

I found in the MNHN at the Library of the French Entomological Society an example of Voet's book where the first pages are missing but *Cerambyx biserratus* was already described in 1778. Since the name *equestris* has been used for over a century I think it is better to conserve the well known name. On the other hand, *Lissonotus shepherdii* is exactly similar to Fabricius' type. To summarize my observation, the most common form with the red band complete seems to be a variety of *L. equestris*. I obtained this abundant species from several legume trees and vines. Both forms emerged from the same logs (interrupted or complete red band). Though the type locality is Cayenne the interrupted form is rarer in French Guiana.

Observed host plants: *Hymenaea courbaril* Linné (Caesalpiniaceae) [Sabatier and Prévost 3490]; *Bauhinia guianensis* Aublet (Caesalpiniaceae) [Scott Mori and al. 23641, 23678]; *Macrolobium sp.* (Caesalpiniaceae) [Scott Mori and al. 23467, 23409].

### Tribe Trachyderini

#### Subtribe Bothriospilina

The identity of *Lamia spinifera* Fabricius, 1792 is not yet resolved. Martins and Moure (1973: 79), giving an illustration of the species, came to the conclusion that it might certainly be an Oriental or Ethiopian species, maybe belonging to the genus *Plocaederus* Thomson.

#### *Knulliana cincta spinifera* (Fabricius, 1792) new combination

*Lamia spinifera* Fabricius, 1792: 275; Fabricius, 1801: 292; Aurivillius, 1923: 606 [Catalog]; Blackwelder, 1946: 627 [Catalog] *incertae sedis*; Zimsen, 1964: 169 [Types]; Martins and Moure, 1973: 79 figs 1-2 (male).  
*Chion ochraceus* Bates, 1885: 243, **new synonymy**  
*Chion cinctus* var. *ochraceus* Schaeffer, 1908: 331  
*Chion cinctum v. ochraceum* Blackwelder, 1946: 562 [Catalog]  
*Cerasphorus cinctum ochraceum* Cazier and Lacey, 1952: 10 fig. 1  
*Knulliana cincta ochracea* Linsley, 1962: 111 fig. 35 (distribution map)

#### Subtribe Ancylocerina

#### *Callancyla croceicollis* (White, 1855)

*Eriphus croceicollis* White, 1855: 292; Bates, 1870b: 429.  
*Calloppisma ruficollis* Bates, 1870b: 419, **new synonymy**  
*Ancylocera ruficollis* Gemminger and Harold, 1872: 2959 [Catalog]  
*Callancyla ruficollis* Aurivillius, 1912: 446 [Catalog]

Bates wrote (1870: 429), concerning *Eriphus croceicollis* White, 1855, "Although taken by me, I do not find the species among my own reserved collection of Amazonian Longicorns". He would never have been able to guess White's misplacement whose type (BMNH) is a female. Bates, though uncertain about the sex of his *Calloppisma ruficollis* supposed the type (MNHN) to be a male.

### Subfamily Lamiinae

#### Tribe Lamiini

#### *Taeniotes farinosus* (Linné, 1758)

*Cerambyx farinosus* Linné, 1758: 390  
*Cerambyx farinosus* DeGeer, 1775: 108, pl. XIV, fig. 1 [Suriname]  
*Cerambyx pulverulentus* Olivier, 1790: 302, **new synonymy**

- Cerambyx surinamensis maculosus* Panzer, 1794: 15, pl. V, fig. 8 [Suriname]  
*Cerambix pulverulentus* Olivier, 1795: 50, (n° 67) pl. VII, fig. 46 b  
*Stenochorus pulverulentus* Schönherr, 1817: 407 [Catalogue, Suriname]  
*Monochamus pulverulentus* Dejean, 1821: 106  
*Taeniotes pulverulentus* Audinet-Serville, 1835: 91 [Cayenne]  
*Monochamus (Taeniotes) farinosus* Castelnau, 1840: 479 [Brazil]  
*Monochamus (Taeniotes) pulverulentus* Castelnau, 1840: 479 [Brazil]  
*Taeniotes farinosus* Bates, 1865b: 110 [Brazil: Amazon, Tapajos]  
*Taeniotes guttularis* Schwarzer, 1929: 364, fig. 16-17  
*Taeniotes pulverulentus m. guttularis* Breuning, 1943b: 248  
*Taeniotes pulvurentus* Bosq, 1943: 109  
*Taeniotes farinosus* Dillon and Dillon, 1941 pl. I, fig. 6  
*Taeniotes farinosa* Blackwelder, 1946: 594 [Catalog]  
*Taeniotes pulverulenta* Blackwelder, 1946: 594 [Catalog]

Olivier's drawing representing *Cerambyx pulverulentus* closely resembles the species commonly collected in Cayenne. Linné's *farinosus*, which is neither in the London Linnean Society nor in Gustave-Adolphe's collection in Uppsala, has in fact no type. Linné mentioned after his laconic description the corresponding illustration in Sybille Merian's book published in 1719 (table 24, f. *infima*) and another reference published by Degeer before the tenth edition of Linné's *Systema Naturae*. The iconotype belongs to the prelinnean literature; I ignore what is to be done in this case. I consulted this very rare book with colored plates and I am certain that *C. pulverulentus* is a synonym of *C. farinosus* and their localities (Suriname and Cayenne) confirm the synonymy. The species is very common on the introduced breadfruit tree *Artocarpus altilis* (S. Parkinson) Fosberg (Moraceae). I do not understand why these two taxa were considered as different by all authors, except for Bates who had already reported the synonymy (1865b: 110).

#### Type Apomecynini

##### *Bisaltes buqueti* Thomson, 1868

- Bisaltes buquetii* Thomson, 1868b: 111 [Cayenne]  
*Bisaltes buqueti* Gemminger and Harold, 1873: 3102 [Catalog]

- Bisaltes sp.* Bates, 1885: 346, pl. IX, fig. 21a  
*Bisaltes batesi* Aurivillius, 1923: 609 [Catalog]  
*Ptericoptus forsteri* Tippmann, 1960: 152, pl. IX, fig. 21a, new synonymy  
*Bisaltes (Bisaltes) buqueti* Breuning, 1960: 177 [Catalog]  
*Bisaltes (s.str.) forsteri* Breuning, 1971: 284

Tippmann's type is in the USNM and the type of Thomson in the MNHN. The species was reared on a Convolvulaceae vine belonging to the genus *Maripa* [Scott Mori *et al.* 23592].

##### *Tethystola brasiliensis* Breuning, 1940

- Tethystola brasiliensis* Breuning, 1940: 42  
*Tethystola flavoapicalis* Breuning, 1942: 141, new synonymy

Both types are at the MNHN and were collected in the Brazilian state of Pernambuco. The type of *T. flavoapicalis* is a big specimen.

#### Tribe Pteropliini

##### *Ataxia obscura* (Fabricius, 1801)

- Stenocorus obscurus* Fabricius, 1801: 307 [America meridionalis]  
*Esthlogena sulcata* Bates, 1866: 290  
*Parysatis collaris* Thomson, 1868: 119  
*Parysatis sulcata* Aurivillius, 1922: 293 (Catalog)  
*Parysatis obscura* Aurivillius, 1922: 292 (Catalog)  
*Ataxia flaviceps* Breuning, 1942: 137, new synonymy  
*Ataxia obscura* Breuning, 1961: 53, fig. 7

##### *Cryptocranium cazieri* Lane, 1958

- Cryptocranium cazieri* Lane, 1958: 6, fig. 2  
*Ornatodesisa pulchra* Breuning, 1961: 19  
*Cryptocranium gounellei* Breuning, 1980: 68, new synonymy

Lane's type (a female from Peru) belongs to the AMNH and Breuning's type (a male with flattened legs from the Brazilian Goiás) belongs to the MNHN. The recent synonymy published by Monné and Giesbert (1992: 252) for a specimen already described by Breuning from the Brazilian state of Pará, and the presence of this species in French Guiana, lead us to consider that this very rare species is widely distributed in northern South America.

##### *Epectasis juncea* (Newman, 1840)

- Saperda juncea* Newman, 1840: 13  
*Epectasis attenuata* Bates, 1866: 294  
*Epectasis grossepunctata* Breuning, 1942: 175, new synonymy

*Epectasis juncea m. attenuata* Breuning, 1961c: 25

Breuning's type belongs to the USNM, the type of *E. attenuata* is in the MNHN.

### Tribe Onciderini

*Paraclytemnestra lineata* (Fisher, 1926), **new combination**.

*Jamesia lineata* Fisher, 1926: 14

*Paraclytemnestra gigantea* Breuning, 1974a: 240, **new synonymy**

This incredible species from the Antilles merits its own genus. Breuning's type belongs to the MNHU, though Fisher's type belongs to the USNM.

### Tribe Agapanthiini

*Hippopsis meinerti* Aurivillius, 1900

*Hippopsis meinerti* Aurivillius, 1900: 416; Aurivillius, 1923: 357 [Catalog]

*Hippopsis meinerti* Blackwelder, 1946: 606 [Catalog]; Galileo and Martins, 1988a: 185 figs 17-20; Galileo and Martins, 1988b: 205

*Hippopsis freyi* Breuning, 1955: 661

*Hippopsis (Hippopsis) lemniscata m. meinerti* Breuning, 1961b: 197 [Catalog]; Breuning, 1962: 10

*Hippopsis (Hippopsis) freyi* Breuning, 1961b: 197 [Catalog]; 1962: 18

*Hippopsis (Hippopsis) lemniscata tobagoensis* Breuning, 1962: 10

*Hippopsis tremata* Galileo and Martins, 1988a: 184, figs. 13-16, **new synonymy**

Aurivillius' type, in the NRS is a male, and the abdominal depression is absolutely identical to the drawing given in the paper of Galileo and Martins to characterize their new species *Hippopsis tremata*. I compared Aurivillius' type with my own material of both sexes from French Guiana and identified by Martins as *H. tremata*.

### Tribe Acanthoderini

*Acanthoderes daviesi* (Swederus, 1787)

*Cerambyx (Lamia) daviesii* Swederus, 1787: 195, pl. VIII, fig. 6

*Cerambyx daviesii* Gmelin, 1790: 1837

*Lamia punctata* Fabricius, 1792: 272 [Cajennae], **new synonymy**

*Lamia daviesii* Schönherr, 1817: 380 [America, Cayenne]

*Acanthoderes daviesii* Dejean, 1835: 336

*Acanthoderes daviesii* Audinet-Serville, 1835: 29 [Cayenne]

*Acanthoderes swederi* White, 1855: 360, pl. IX, fig. 6 [Para]

*Acanthoderes daviesi* Thomson, 1864: 17

*Acanthoderes daviesi* var. *swederi* Gemminger and Harold, 1873: 3146 [Catalog]

*Acanthoderes daviesi* Bodkin, 1919: 268

*Acanthoderes (Acanthoderes) daviesi* Aurivillius, 1923: 384 [Catalog]

*Acanthoderes (Acanthoderes) punctata* Aurivillius, 1923: 385 [Catalog]

*Acanthoderes (Acanthoderes) daviesi* Buck, 1959: 604

The common and widely distributed epiphyte *Clusia grandiflora* Splitgerber (Clusiaceae) [Denis Loubry 1827] breeds *A. daviesi*.

### Tribe Acanthocinini

*Oedopeza ocellator* (Fabricius, 1801), **new combination**

*Lamia ocellator* Fabricius, 1801: 287 [America meridionali]

*Leiopus (Edopeza) pogonocheroides* Audinet-Serville, 1835: 88, **new synonymy**

*Leiopus pogonocheroides* Castelnau, 1840: 464 [Cayenne]

*Oedopeza pogonocheroides* White, 1855: 394

*Ozineus ocellator* Aurivillius, 1923: 398 [Catalog]

I do not understand why this very common species was not recognized by Aurivillius. The type of Fabricius is a male with a characteristic swollen protarsal segment. I obtained this species from 16 different tree species, mainly belonging to the legume group: *Couma guianensis* Aublet (Apocynaceae) [Christian Feuillet 2311]; *Simarouba amara* Aublet (Simaroubaceae) [Christian Feuillet 2375]; *Theobroma cacao* Linné (Sterculiaceae) [Jean-Jacques de Granville det.]; *Swartzia lomatophylla* (Benth) Pittier (Caesalpiniaceae) [Denis Loubry, 1585]; *Swartzia panacoco* (Aublet) Cowan (Caesalpiniaceae) [Denis Loubry, 1804]; *Machaerium* sp. (Papilionaceae) [Denis Loubry, 1815]; *Taralea oppositifolia* Aublet (Papilionaceae) [Denis Loubry, 1611]; *Abarema curvicarpa* (Irwin) Barneby and Grimes (Mimosaceae) [Grimes and Sabatier 3312]; *Abarema jupunba* (Willdenow) Barneby and Grimes (Mimosaceae) [Christian Feuillet 2307, Denis Loubry 1875 and Scott Mori 23688]; *Balizia pedicellaris* (de Candolle) Barneby and Grimes (Mimosaceae) [Christian Feuillet 1341]; *Enterolobium schomburgkii* Benth (Mimosaceae) [Daniel Sabatier 2367]; *Enterolobium* sp. (Mimosaceae) [Denis Loubry 1763]; *Hydrochorea corymbosa* (L.C.

Richard) Barneby and Grimes (Mimosaceae) [Denis Loubry 1300]; *Inga melinonis* Sagot (Mimosaceae) [Denis Loubry 1780]; *Inga* sp. (Mimosaceae) [Scott Mori 23456]; *Parkia nitida* Miquel (Mimosaceae) [Daniel Sabatier 1511 and Denis Loubry 1564].

***Toronaeus incisus* (Bates, 1864), new combination.**

*Nyssodryes incisus* Bates, 1864: 155

*Nyssodryes incisus* Gilmour, 1965: 599 [Brazil] Catalogue

*Stenolis incisus* Monné, 1985: 535.

The rearing of this rare species demonstrates clearly its greater affinity with the genus *Toronaeus* (where majority were bound to Burseraceae trees), confirmed by its taxonomic features. It was obtained on a still unidentified Burseraceae [Denis Loubry 1318]

***Xylergates elaineae* Gilmour, 1962**

*Astynomus hilaris*: Prudhomme, 1906: 38

*Xylergates* sp. (*hilaris* Dejean?) Duffy, 1960: 258, figs. 158-159 (Larva)

*Xylergates elaineae* Gilmour, 1962: 277, pl. III, fig. 3

*Xylergates dorotheae* Gilmour, 1962: 279, pl. III, fig. 4, **new synonymy**

Gilmour's two types belong to the American Museum of Natural History in New York. The species commonly attacks fallen trees belonging to the family of Lecythidaceae. I found this species from seven different host plants in French Guiana: *Lecythis poiteaui* O.C. Berg [Scott Mori det. 1995]; *Lecythis holcogyne* (Sandwith) Mori [D. Sabatier and M.F. Prévost 2108]; *Eschweilera sagotiana* Miers [Daniel Sabatier 1762]; *Eschweilera micrantha* (O.C. Berg) Miers [Scott Mori 23479]; *Eschweilera coriacea* (A.P. de Candolle) Martius ex O.C. Berg [Denis Loubry 1846, 1341]; *Eschweilera congestiflora* (R. Benoist) Eyma [Scott Mori 23478]; *Couratari guianensis* Aublet [Daniel Sabatier 1771 and Denis Loubry 1337].

**Tribe Colobotheini**

***Colobothea macularis* (Olivier, 1792)**

*Lamia macularis* Olivier, 1792: 474 [Suriname]

*Cerambyx macularis* Olivier, 1795: (67) 98 pl. XX, fig. 154 [Suriname]

*Colobothea velutina* Bates, 1865a: 216, **new synonymy**

*Colobothea macularis*: Aurivillius, 1923: 454 [Catalog]

I had the opportunity to consult and to photograph the original paintings of Olivier which served to illustrate his famous "Entomologie..." still conserved in his family, six generations later. These paintings reveal more details than the printed and post-colored plates found in the remaining examples of this very rare book and thus permit the above synonymy. I obtained several adults from *Hydrochorea corymbosa* (L.C. Richard) Barneby and Grimes (Mimosaceae) [Denis Loubry 1300]

**Tribe Pretiliini**

***Pretilia tuberculata* (Fabricius, 1801), new combination.**

*Saperda tuberculata* Fabricius, 1801: 331 [America meridionali]

*Pretilia telephoroides* Bates, 1866: 302; Lacordaire, 1872: 910; Gilmour, 1962: 125; Martins and Galileo, 1990b: 707 figs 1-11, **new synonymy**

*Eupogonius tuberculatus* Aurivillius, 1923: 312 [Catalog]; Blackwelder, 1946: 600 [Catalog]

**Tribe Calliini**

***Drycothaea angustifrons* (Breuning, 1943)**

*Estoloides (Estoloides) angustifrons* Breuning, 1943a: 56 [Guyane: Maroni]; Breuning, 1974b: 59.

*Guyanestola macrophthalma* Breuning, 1961a: 248, **new synonymy**

*Guyanestola macrophthalma* Breuning, 1963: 507 [Catalog]

*Drycothaea angustifrons* Tavakilian, 1991: 450; Galileo and Martins, 1991: 250

*Drycothaea macrophthalma* Galileo and Martins, 1991: 251

In 1990, the blocked situation concerning Frey's collection made impossible the examination of Breuning's type during my journey to Munich. The situation is now normal since the collection was transferred from Frey's house in Tutzing to the Staatliches Museum in Munich (ZSMA). I am grateful to Patrick Bleuzen who sent me the picture of *Guyanestola macrophthalma* and checked the claws, confirming that they are appendiculated as Galileo and Martins and I already guessed. This illustrates how uncomfortable can be the position of an entomologist revising a group of which the types are still in private collections.

I obtained this species from *Cecropia palmata* Willdenow (Cecropiaceae) [Denis Loubry 1570].

***Drycothaea brasiliensis* (Breuning, 1974)**

*Guyanestola brasiliensis* Breuning, 1974b: 49 [Brazil: Amazonas]

*Drycothaea marmorata* Martins and Galileo, 1990a: 611, fig. 4, **new synonymy**

*Drycothaea brasiliensis* Galileo and Martins, 1991: 251

Breuning's type is in the ZSMA. The host plant is still *Cecropia palmata* Willdenow (Cecropiaceae) [Denis Loubry 1570].

***Drycothaea truncatipennis* new name for *Estola stictica* Breuning, 1942 (nec Bates, 1881)**

*Estola stictica* Breuning, 1942: 163

The type belongs to the USNM (ex Tippmann's collection n° 41605) and is a male collected in the Brazilian state of Santa Catarina. The structure of the claws transfers this species from the tribe Desmiphorini to Calliini in the genus *Drycothaea*. Unfortunately another *Drycothaea stictica* Bates, 1881 already exists from Guatemala. I propose the specific name *D. truncatipennis* which fits better with the elytral aspect of that misplaced species.

***Drycothaea turrialbae* (Breuning, 1943), new combination**

*Estoloides (Estoloides) turrialbae* Breuning, 1943: 56

*Estoloides (Estoloides) turrialbae* Breuning, 1963: 507

*Estoloides turrialbae* Chemsak, Linsley and Noguera, 1992: 119 (Catalog)

The type belongs to the USNM (ex Tippmann's collection n° 41596) and is a female labelled as *Parestola turrialbae*! As seen above, this species is transferred from Desmiphorini to the tribe Caliini for the same reason.

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### Book Review

*Biologie des Coleopteres Chrysomelides*. Pierre Jolivet. Boubee Publisher, Paris, 1997, 280 pp. (USA \$57.00)

The main biological aspects of this economically important group of beetles, the Chrysomelidae, are summarized in this new work by the internationally known author, Dr. Pierre Jolivet. Around 37,000 species are now described and more remain to be named. The size and diversity of the family are less than those of the weevils (Curculionidae), but probably there will be a total of 40 to 50 thousand species once the canopy fauna is better known. For instance, we still know nothing about the biology and the host plants of the Madagascar leaf beetles and surely many more species from that region await description.

The importance of the group is mostly agricultural as might be expected from a group almost entirely leaf, stem, and root feeders. Members range in length from 1 mm, or less, up to 27 mm. Many species are brightly colored.

Probably the family is polyphyletic, *i.e.*, composed of morphologically similar but phylogenetically distinct groups, and with Bruchidae as its sister group, all have Cerambycoid ancestors. Unquestioned chrysomelid fossils date from the Jurassic.

The book covers the palaeontology, evolution, development, and adaptations of the family. Mimicry, defensive reactions, pathogens and parasites, and every aspect of the biology and ecology of the beetles are discussed.

The classification adopted for this book is not that of cladists. Jolivet recognizes 20 extant subfamilies, and one extinct subfamily, probably the direct Jurassic ancestor of the Aulacoscelinae. The larvae of several subfamilies were still unknown at the time this book was written, such as those of the Megascelinae and Aulacoscelinae, but recently, I am told by the author, these larvae have been obtained and are now being described.

The author believes that the recently proposed monophyletic classifications do not reflect the real relationships within the family. He disagrees with the fusion of Alticinae and Galerucinae, of Hispinae and Cassidinae, and Synetinae and Megascelinae, included with the Eumolpinae, as well as the elevation to family rank of some subfamilies. *Syneta*, for instance, has its wings and male genitalia completely different from those of the Eumolpinae. In the book he argues strongly that the relationships recognized within the Chrysomelidae be based on their food-plant selection when these are known, wing venation, and male genitalia, and use both physiological and mechanical characters of the plants.

Line drawings illustrate the book and six colored plates show the beetles in various tropical settings in both the Old and New World. Three other plates include some water color drawings of spectacular species.

The author, with several collaborators, has written five specialized books on the biology of Chrysomelidae prior to this volume. Here, the author presents a new synthesis of the topic, including many aspects not found in the previous volumes. The collected works were noted on pages 260 and 316 of volume 10, 1996, of *Insecta Mundi*.

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